

TRITERPENOID AND PHENOLIC COMPOUNDS FROM *EUPHORBIA TITHYMALOIDES* L. (EUPHORBIACEAE)

Vu Minh Trang, Pham Thu Hien, Vu Hoang Nam, Phan Tong Son, Phan Minh Giang*

Faculty of Chemistry, VNU University of Science, Vietnam National University, Hanoi

Received 26 January 2014

Abstract

Friedelan-3 β -ol (**1**), β -sitosterol (**2**), oryzanol-C (**3**), 2,4,6-trimethoxyacetophenone (**4**), 3,4,3'-tri-*O*-methylellagic acid (**5**), scoparone (**6**), hopenone B (**7**), and tetradecane-1,2-diol (**8**) were isolated from the leaves and stems of *Euphorbia tithymaloides* L. (Euphorbiaceae) collected in northern Vietnam. Their structures were determined by spectroscopic analysis.

Keywords: *Euphorbia tithymaloides*, *Pedilanthus tithymaloides*, Euphorbiaceae, triterpenoid, phenolic compound.

1. INTRODUCTION

Euphorbia is the largest genus in the spurge (Euphorbiaceae) family and an interesting genus for natural product chemists owing to its biological diversity, large distribution, diverse chemistry, and specific anticancer and multidrug-resistant activities of its constituents [1]. *Euphorbia tithymaloides* L. (syn. *Pedilanthus tithymaloides* (L.) Poit.) is a perennial succulent spurge. The shrub is native to tropical and North America and Central America. The Vietnamese species (local name: *Thuoc dau*) is usually grown as medicinal and ornamental plant [2]. Its fresh leaves are used to treat cuts and wounds. A few phytochemical investigations on *P. tithymaloides* reported the isolation of long-chain alcohols, sterols, terpenes [3, 4], and flavonoids [5], including a noticeable report on antiplasmodial and antimycobacterial poly-*O*-acylated jatrophane diterpenes from the white latex [3].

2. EXPERIMENTAL

2.1. General

ESI-MS spectra were measured on a LC-MS Agilent 6310 system. ^1H -NMR, ^{13}C -NMR, and DEPT spectra were recorded on a Bruker Avance 500 NMR spectrometer. Diaion HP-20 (Mitsubishi, Japan) and silica gel Merck (Darmstadt, Germany) were used for column chromatography (CC). Thin-layer chromatography (TLC) was performed on precoated silica gel Merck 60 F₂₅₄ plate.

2.2. Plant Materials

The fresh leaves and stems of *E. tithymaloides* were collected in Hanoi, Vietnam two times; the first collection (14 kg) in April 2009 and the second collection (15 kg) in June 2010. A voucher specimen (HCTN 409) has been deposited in the Laboratory of Chemistry of Natural Products, VNU University of Science, Vietnam National University, Hanoi.

2.3. Extraction and Isolation of 1-8

The samples were air-dried and then oven-dried at 45–50 °C. The materials for the extraction were 1.3 kg of the dried stems and 334.7 g of the dried leaves from the first collection and 1 kg of the dried stems and 3.5 kg of the fresh stems from the second collection. The general extraction procedure involved maceration of the samples with MeOH at room temperature, followed by fractionation of the resultant MeOH extract into *n*-hexane-, CH₂Cl₂-, and EtOAc-soluble fractions by liquid-liquid extraction between water and the organic solvents in the order of the increasing polarities. The *n*-hexane-soluble fractions from the leaves (10 g) and stems (14 g) of the first collection were separated by repeated silica gel column chromatography using gradient *n*-hexane-acetone as the mobile phase and then further recrystallization gave **1** (18.6 mg), **2** (100 mg), and **3** (30 mg). Similar TLC patterns were observed for the *n*-hexane- and CH₂Cl₂-soluble fractions from the fresh stems (3.4 g and 11 g, respectively) as well as the dried stems (5.2 g and 6 g, respectively); they were combined accordingly and subjected to

repeated silica gel column chromatography using gradient *n*-hexane-EtOAc to give **4** (36.1 mg), **5** (5 mg), **6** (76.5 mg), and **7** (2 mg). The water phase from the extraction of the dried stems was concentrated and chromatographed on Diaion HP-20 using MeOH-H₂O 20 %, 40 %, and 60 %. The fraction eluted with MeOH-H₂O 60% was chromatographed on silica gel eluting with gradient CH₂Cl₂-MeOH 19:1, 9:1, and 6:1 to give **8** (10 mg).

Friedelan-3 β -ol (1): White needles, m.p. 281–282 °C. ¹H-NMR (CDCl₃, δ , ppm, *J*/Hz): 0.86 (3H, s, CH₃-25), 0.94 (3H, d, *J* = 7.5 Hz, CH₃-23), 0.95 (3H, s, CH₃-24), 0.97 (3H, s, CH₃-30), 0.99 (6H, s, CH₃-26, CH₃-27), 1.01 (3H, s, CH₃-29), 1.17 (3H, s, CH₃-28), 3.73 (1H, s, H-3). ¹³C-NMR/DEPT (CDCl₃): 11.6 (q, C-23), 15.8 (t, C-7), 16.4 (q, C-24), 17.6 (t, C-1), 18.3 (q, C-25), 18.6 (q, C-27), 20.1 (q, C-26), 28.2 (s, C-20), 30.1 (s, C-17), 30.7 (t, C-12), 31.8 (q, C-30), 32.1 (q, C-28), 32.4 (t, C-15), 32.9 (t, C-21), 35.0 (q, C-29), 35.2 (t, C-2), 35.4 (t, C-19), 35.6 (t, C-11), 36.1 (t, C-16), 37.1 (s, C-5), 37.9 (s, C-9), 38.4 (s, C-14), 39.3 (t, C-22), 39.7 (s, C-13), 41.8 (t, C-6), 42.9 (d, C-18), 49.2 (d, C-4), 53.2 (d, C-8), 61.4 (d, C-10), 72.8 (d, C-3).

β -Sitosterol (2): White needles, m.p. 134–136 °C. IR (KBr): ν_{max} cm⁻¹ 3427, 1637, 1461, 1379, 1056.

Oryzanol-C (3): White needles, m.p. 198–199 °C. ESI-MS (positive mode): *m/z* 639.6 ([M+Na]⁺, C₄₁H₆₀O₄Na). ¹H-NMR (CDCl₃, δ , ppm, *J*/Hz): 0.18 (1H, d, *J* = 3.9 Hz, H-19a), 0.43 (1H, d, *J* = 3.9 Hz, H-19b), 0.89 (3H, s, CH₃-28), 0.89 (3H, d, *J* = 6.4 Hz, CH₃-21), 0.91 (3H, s, CH₃-30), 0.92 (3H, s, CH₃-29), 0.98 (3H, s, CH₃-18), 1.03 (3H, d, *J* = 6.8 Hz, CH₃-26), 1.04 (3H, d, *J* = 6.8 Hz, CH₃-27), 2.24 (1H, septet, *J* = 6.8 Hz, H-25), 3.93 (3H, s, CH₃O-3'), 4.64 (1H, dd, *J* = 10.5 Hz, 4.5 Hz, H-3), 4.65 (1H, br s, H-31a), 4.72 (1H, br s, H-31b), 6.3 (1H, d, *J* = 15.5 Hz, H- α), 6.91 (1H, d, *J* = 8.5 Hz, H-5'), 7.04 (1H, d, *J* = 1.5 Hz, H-2'), 7.07 (1H, dd, *J* = 8.5 Hz, 1.5 Hz, H-6'), 7.61 (1H, d, *J* = 15.5 Hz, H- β).

2,4,6-Trimethoxyacetophenone (4): White amorphous powder. ¹H-NMR (CDCl₃, δ , ppm, *J*/Hz): 2.46 (3H, s, CH₃CO), 3.79 (6H, s, CH₃O-2, CH₃O-6), 3.83 (3H, s, CH₃O-4), 6.1 (2H, s, H-3, H-5).

3,4,3'-Tri-O-methyllellagic acid (5): White amorphous powder. ¹H-NMR (CDCl₃ + CD₃OD, δ , ppm, *J*/Hz): 4.05 (3H, s), 4.22 (3H, s), 4.26 (3H, s) (CH₃O-3, CH₃O-3', CH₃O-4), 7.68 (1H, s), 7.69 (1H, s) (H-5, H-5').

Scoparone (6): White amorphous powder. ¹H-NMR (CDCl₃, δ , ppm, *J*/Hz): 3.93 (3H, s, CH₃O-7), 3.96 (3H, s, CH₃O-6), 6.29 (1H, d, *J* = 9.5 Hz, H-3), 6.84 (1H, s, H-8), 6.86 (1H, s, H-5), 7.62 (1H, d, *J* =

9.5 Hz, H-4).

Hopenone B (7): White amorphous powder. ¹H-NMR (CDCl₃, δ , ppm, *J*/Hz): 0.73 (3H, s, CH₃-28), 0.93 (3H, s, CH₃-25), 0.95 (3H, s, CH₃-27), 1.01 (1H, s, CH₃-26), 1.03 (3H, s, CH₃-23), 1.07 (3H, s, CH₃-24), 1.75 (3H, s, CH₃-30), 2.4 (1H, ddd, *J* = 15.5 Hz, 8.0 Hz, 7.5 Hz, H-2a), 2.5 (1H, ddd, *J* = 15.5 Hz, 7.5 Hz, 4.0 Hz, H-2b), 2.69 (1H, br q, *J* = 6.5 Hz, H-21), 4.78 (2H, br s, 2H-29). ¹³C-NMR/DEPT (CDCl₃): 15.7 (q, C-25), 16.2 (q, C-28), 16.4 (q, C-26), 16.6 (q, C-27), 19.8 (t, C-6), 21.1 (q, C-24), 21.59 (t, C-11), 21.6 (t, C-16), 23.9 (t, C-12), 24.9 (q, C-30), 26.6 (q, C-23), 27.4 (t, C-20), 32.7 (t, C-7), 33.7 (t, C-15), 34.2 (t, C-2), 36.9 (s, C-10), 39.6 (t, C-1), 41.7 (s, C-8), 41.9 (t, C-19), 42.2 (s, C-14), 44.8 (s, C-18), 46.5 (d, C-21), 47.4 (s, C-4), 49.6 (d, C-13), 49.7 (d, C-9), 54.9 (2 \times d, C-5, C-17), 110.2 (t, C-29), 148.6 (s, C-22), 218.1 (s, C-3).

Tetradecane-1,2-diol (8): White amorphous powder. ESI-MS (positive mode): *m/z* 222 ([M+Na-CH₂OH]⁺, C₁₄H₃₀O₂Na-CH₂OH). ¹H-NMR (CD₃OD, δ , ppm, *J*/Hz): 0.92 (3H, t, *J* = 7.0 Hz, CH₃-14), 1.31 (20H, br s, CH₂-4 → CH₂-13), 1.54 (2H, m, CH₂-3), 3.79 (1H, m, H-2), 3.9 (1H, dd, *J* = 10.5 Hz, 6.0 Hz, H-1a), 3.96 (1H, dd, *J* = 10.5 Hz, 4.5 Hz, H-1b).

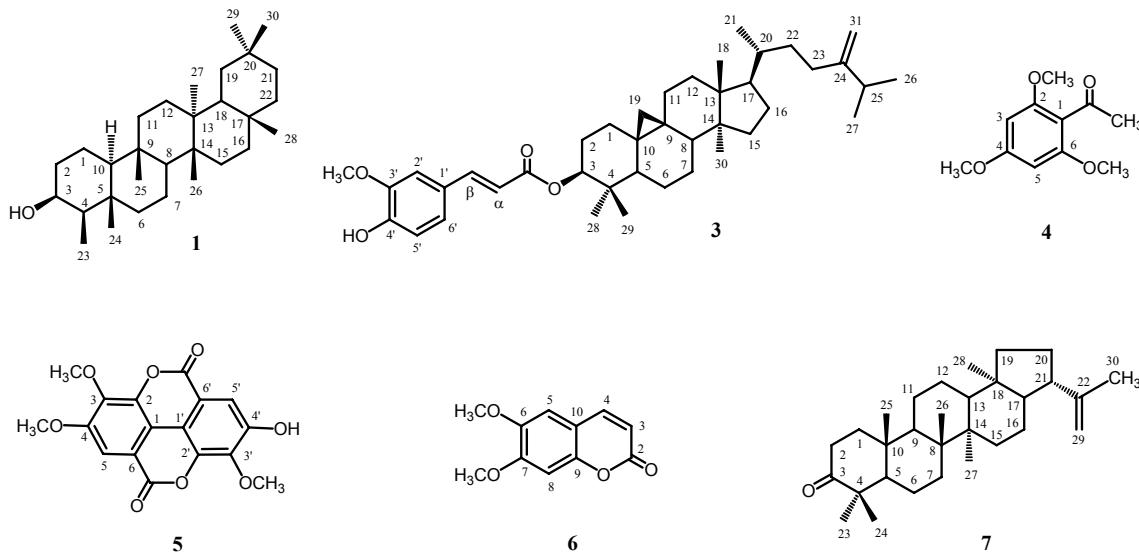
3. RESULTS AND DISCUSSION

The leaves and stems of *E. tithymaloides* were extracted with MeOH and the extracts were partitioned between water and organic solvents. The *n*-hexane- and CH₂Cl₂-soluble fractions were chromatographed repeatedly on silica gel to yield seven compounds. The following triterpenoids and phenolic compounds were isolated for the first time from *E. tithymaloides*, friedelan-3 β -ol (**1**) [3, 6], oryzanol-C (**3**) [7], 2,4,6-trimethoxyacetophenone (**4**) [8], and hopenone B (**7**) [9]. β -Sitosterol (**2**), 3,4,3'-tri-O-methyllellagic acid (**5**) [10], and scoparone (**6**) [11] isolated from the *n*-hexane- and CH₂Cl₂-soluble fraction of the leaves and stems; and tetradecane-1,2-diol (**8**) [12] from the water phase of the dried stems are the known compounds from *E. (Pedilanthus) tithymaloides* [3, 4, 13]. The structures of compounds **1–8** were determined by comparing their MS and NMR spectroscopic data with those reported in the literature [6–11].

Compound **1** was isolated from the *n*-hexane-soluble fraction as white needles, m.p. 281–282 °C. The ¹H-NMR (CDCl₃) spectrum of **1** implied the presence of seven singlet methyl groups [δ_H 0.86 (3H, s), 0.95 (3H, s), 0.97 (3H, s), 0.99 (6H, s), 1.01 (3H, s), and 1.17 (3H, s)], a secondary methyl group [δ_H 0.94 (3H, d, *J* = 7.5 Hz)], and an oxymethylene [δ_H

3.73 (1H, s)]. The ^{13}C -NMR (CDCl_3) and DEPT spectra of **1** showed thirty carbon-13 signals including eight methyl groups (8q) at δ_{C} 11.6, 16.4, 18.3, 18.6, 20.1, 31.8, 32.1, and 35.0; eleven methylenes (11t) at δ_{C} 15.8, 17.6, 30.7, 32.4, 32.9, 35.2, 35.4, 35.6, 36.1, 39.3, and 41.8; five methines

(5d) at δ_{C} 42.9, 49.2, 53.2, 61.4, and 72.8; and six quaternary carbons (7s) at δ_{C} 28.2, 30.1, 37.1, 37.9, 38.4, and 39.7. The ^1H -NMR and ^{13}C -NMR spectra determined the structure of **1** to be friedelan-3 β -ol [3, 6].



Compound **3** was isolated from the *n*-hexane-soluble fraction as white needles, m.p. 198–199 °C. The positive-mode ESI-MS of **3** gave a quasi-molecular ion peak at m/z 639.6 ($[\text{M}+\text{Na}]^+$), indicating a molecular formula $\text{C}_{41}\text{H}_{60}\text{O}_4$. The ^1H -NMR (CDCl_3) spectrum of **3** identified a triterpenoid of 24-methylenecycloartane series. The signals for the cyclopropane methylene at δ_{H} 0.18 (1H, d, $J = 3.9$ Hz) and 0.43 (1H, d, $J = 3.9$ Hz); four tertiary methyl groups at δ_{H} 0.89 (3H, s), 0.92 (6H, s), and 0.98 (3H, s); a secondary methyl group at δ_{H} 0.89 (3H, d, $J = 6.3$ Hz); and an isopropyl group at δ_{H} 1.03 (3H, d, $J = 6.8$ Hz) and 1.04 (3H, d, $J = 6.8$ Hz) were observed. In the downfield region signals for the double bond methylene at δ_{H} 4.65 (1H, s) and 4.72 (1H, s) and an oxymethylene at δ_{H} 4.64 (1H, dd, $J = 10.5$ Hz, 4.5 Hz) were observed. A feruloyl group [δ_{H} 6.91 (1H, d, $J = 8.5$ Hz), 7.04 (1H, d, $J = 1.5$ Hz), and 7.07 (1H, dd, $J = 8.5$ Hz, 1.5 Hz); 6.3 (1H, d, $J = 15.5$ Hz) and 7.61 (1H, d, $J = 15.5$ Hz); and 3.93 (3H, s)] was bonded to C-3 causing downfield shift of H-3 (δ_{H} 4.64) in comparison of that of 24-methylenecycloartanol ($\delta_{\text{H},3}$ 3.28 (CDCl_3); $\Delta\delta_{\text{H}}$ +1.36 ppm) [4]. This feruloyl group was determined to be β -oriented on the basis of the proton-proton coupling constant between H-2_{ax} and H-3_{ax} ($J = 10.5$ Hz). On the basis of the NMR analysis, **3** was determined to be 24-methylenecycloartanyl ferulate (oryzanol-C) [5].

Compound **4** was isolated from the *n*-hexane-soluble fraction as a white amorphous powder. Inspection of the ^1H -NMR spectrum of **4** exhibited the presence of an acetyl group at δ_{H} 2.46 (3H, s), three methoxy groups at δ_{H} 3.79 (6H, s) and 3.83 (3H, s), and a two-proton singlet of an substituted benzene at δ_{H} 6.10 (2H, s). The ^1H -NMR spectroscopic data indicated that **4** possessed an acetophenone-type structure and its substitution pattern was either 2,4,6-trimethoxy [8] or 3,4,5-trimethoxy [14]. The ^1H -NMR data of **4** were superimposed on those of 2,4,6-trimethoxyacetophenone.

Compound **7** was isolated from the *n*-hexane-soluble fraction as a white amorphous powder. In the ^1H -NMR spectrum of **7**, six methyl groups resonated at δ_{H} 0.73 (3H, s), 0.93 (3H, s), 0.95 (3H, s), 1.01 (3H, s), 1.03 (3H, s), and 1.07 (3H, s), and an isopropenyl group at δ_{H} 1.75 (3H, s) and 4.78 (2H, s br). Thirty ^{13}C -NMR signals including seven methyl groups, ten methylenes, five methines, five carbons, a carbonyl ketone [δ_{C} 218.1 (s, C-3)], and a 1,1-disubstituted double bond [δ_{C} 110.2 (t, C-29) and 148.6 (s, C-22)] suggested a triterpenoid structure of **7**. The presence of the isopropylene implied the lup-22(29)-ene or hop-22(29)-ene structure of **7**. By comparing the ^{13}C -NMR spectroscopic data of **7** with those of literature values the structural similarity of **7** and 2-oxohopan-22-ol (rings A, B,

and C) and hop-22(29)-en (rings D and E) was seen [15]. On the basis of the NMR analysis 7 was determined to be 3-oxo-hop-22(29)-en; this structure occurs in nature in the form of two C-21 epimers: moretenone (21α -H) [16, 17] and hopenone B (21β -H) [9]. The stereochemistry at C-21 greatly affected the chemical shifts of C-21 and C-30: δ_C 47.9 (C-21) and 19.7 (C-30) of mortenone and 46.5 (C-21) and 25.0 (C-30) of hop-22(29)-ene [15]. The latter values agreed well with those of 7. Therefore, the structure of 7 was determined to be hopenone B. Hopenone B is a rare naturally occurring compound which was isolated from *Euphorbia cyparissias* in 1969 and structurally elucidated with insufficient $^1\text{H-NMR}$ data [9].

4. CONCLUSION

The study isolated eight compounds from the leaves and stems of *E. tithymaloides*. Of the compounds isolated triterpenoids and phenolic compounds are of special interest. Friedelane, cycloartane, and hopane triterpenoids may be biogenetically related in this plant.

Acknowledgement: This research is funded by Vietnam National Foundation for Science and Technology Development (NAFOSTED) under grant number 104.01-2012.10.

REFERENCES

- Q. W. Shi, X. H. Su, H. Kiyota. *Chemical and pharmacological research of the plants in genus Euphorbia*, Chem. Rev., **108**, 4295-4327 (2008).
- Đỗ Tất Lợi. *Những cây thuốc và vị thuốc Việt Nam*, Nhà xuất bản Y học, Hà Nội (2001).
- W. Mongkolvisut, S. Sutthivaiyakit. Antimalarial and antituberculous poly-O-acylated jatrophane diterpenoids from *Pedilanthus tithymaloides*, J. Nat. Prod., **70**, 1434-1438 (2007).
- S. Ghosh, A. Samanta, N. B. Mandal, S. Bannerjee, D. Chattopadhyay. *Evaluation of the wound healing activity of methanol extract of Pedilanthus tithymaloides (L.) Poit leaf and its isolated active constituents in topical formulation*, J. Ethnopharmacol., **142**, 714-722 (2012).
- P. M. Abreu, S. Matthew, T. González, L. Vanickova, D. Costa, A. Gomes, M. A. Segundo, E. Fernandes. *Isolation and identification of antioxidants from Pedilanthus tithymaloides*, J. Nat. Med., **62**, 67-70 (2008).
- S. Monkodaev, C. Laetchutinat, N. Nuntasen, W. Popimon. *Identification and antiproliferative activity evaluation of a series of triterpenoids isolated from Flueggea virosa Roxb. ex. Willd.*, American J. Applied Sciences, **6**, 1800-1806 (2009).
- K. Yasukawa, T. Akihisha, Y. Kimura, T. Tamura, M. Takido. *Inhibitory effect of cycloartenol ferulate, a component of rice bran, on tumor promotion in two-stage carcinogenesis in mouse skin*, Biol. Pharm. Bull., **21**, 1072-1076 (1998).
- N. Ruangrungsi, P. Tantivanata, R. P. Borris, G. A. Cordell. *Traditional medicinal plants of Thailand. III. Constituents of Zanthoxylum budrunga (Rutaceae)*, J. Sci. Soc. Thailand, **7**, 123-127 (1981).
- A. N. Starnatt. *Isolation of hopenone B from Euphorbia cyparissias*, Phytochemistry, **8**, 1831-1832 (1969).
- S. Zhang, X. Liu, Z. L. Zhang, L. He, Z. Wang, G. S. Wang. *Isolation and identification of the phenolic compounds from the roots of Sanguisorba officinalis L. and their antioxidant activities*, Molecules, **17**, 13917-13922 (2012).
- C. H. Ma, W. Ke, Z. L. Sun, J. Y. Peng, Z. X. Li, X. Zhou, G. R. Fan, C. G. Huang. *Large-scale isolation and purification of scoparone from herba Artemisia scopariae by high-speed counter-current chromatography*, Chromatographia, **64**, 83-87 (2006).
- Y. Kinjo, B. Bei, S. Bufali, R. Raju, S. K. Richardson, M. Imamura, M. Fujio, D. Wu, A. Khurara, K. Kawahara, C-H. Wong, A. R. Howell, P. H. Seeberger, M. Kronenberg. *Natural Sphingomonas glycopipids vary greatly their ability to activate natural killer T cells*, Chemistry & Biology, **15**, 654-664 (2008).
- C. T. Inh, N. M. Cuong, N. A. Hung, N. T. Huong, P. Q. Long. *Study of chemical constituents of Euphorbia tithymaloides (P.)*, Vietnam J. Chemistry, **51**, 309-313 (2013).
- T. Yokoyama, H. M. Chang, R. S. Reiner, R. H. Atalla, I. A. Weinstock, J. F. Kadla. *Polyoxometalate oxidation of non-phenolic lignin subunits in water: effect of substrate structure on reaction kinetics*, Holzforschung, **58**, 116-121 (2004).
- Trương Thị Tố Chinh, Lưu Thị Kim Nhhung, Đỗ Thị Kim Huệ, Phan Minh Giang, Phan Tông Sơn. *Các tritecpenoit, flavonoit, diarylheptanoit và các hợp chất thành phần khác từ cây Tống quán sủi (*Alnus nepalensis* D. Don, Betulaceae)*, Tạp chí Hóa học, **49**, 196-202 (2011).
- J. P. David, M. Meira, J. M. David, M. L. da S. Guedes. *Triterpenos e ferulatos de alquila de Maprouna guianensis*, Quim. Nova, **27**, 62-65 (2004).
- G. Degaldo, J. Hernández, M. Y. Kios, M. I. Aguilar. *Pentacyclic triterpenes from Cnidoscolus multilobus*, Planta Med., **60**, 384-390 (1994).

Corresponding author: Phan Minh Giang

Faculty of Chemistry, VNU University of Science, Vietnam National University, HN
19 Le Thanh Tong, Hoan Kiem, Hanoi Vietnam
Email: phanminhgiang@yahoo.com.

